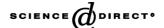


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# Development of an anti-core lipopolysaccharide vaccine for the prevention and treatment of sepsis

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### Abstract

Sepsis continues to be a leading cause of death among hospitalized patients. Despite advances in supportive care and the availability of potent antimicrobials, the mortality exceeds 20%. The passive infusion of antibodies directed against a conserved region of the lipopolysaccharide (LPS) of Gram-negative bacteria was highly protective in an early study (NEJM 307 [1982] 1225). When this and similar preparations were unable to show consistent efficacy, efforts were directed towards other strategies, including cytokine modulation. Our group found that a whole bacterial vaccine made from the Escherichia coli O111:B4, J5 (Rc chemotype) mutant induced protective antibodies when given passively as treatment for sepsis in a neutropenic rat model. A subunit vaccine, composed of detoxified J5 LPS complexed to group B meningococcal outer membrane protein (OMP), provided similar protection when antibodies were given passively, or induced actively in both the neutropenic and cecal ligation/puncture models of sepsis. A phase I study in 24 subjects (at 5, 10 and 25 µg doses [based on LPS] for each group of 8) revealed the vaccine to be well-tolerated with no systemic endotoxin-like effects. Although a two to three-fold increase in antibody levels over baseline (by ELISA assay) was observed at the 10 and 25 µg doses, the plasma from both high and low responders reduced LPS-induced cytokine generation in whole blood. Reimmunization of six subjects at 12 months did not convert low responders to high responders or boost the still elevated anti-J5 LPS levels of high responders. If functional assays of anti-LPS antibodies are better predictors of vaccine efficacy than ELISA antibody levels, then it will be necessary to determine which of many potential assays best correlates with protection in animal models. We are currently comparing a panel of functional assays with protective efficacy in animal models of sepsis, as well as the ability of adjuvants to enhance vaccine efficacy. The availability of an effective anti-endotoxin vaccine will provide additional therapeutic options for the prevention and/or treatment of sepsis. © 2003 Elsevier Ltd. All rights reserved.

Keywords: Sepsis; Vaccine; Anti-endotoxin antibody; Lipopolysaccharide

## 1. Introduction

Sepsis, a leading cause of death in intensive care units, has increased in frequency over the last two decades [1]. Between 1979 and 2000 there was a four-fold increase in the number of cases of sepsis (from 164,000 to nearly 660,000). The mortality remains nearly 20% despite advances in supportive care and the introduction of potent antimicrobial agents [1]. Consequently, additional therapeutic measures have been sought. The important role of Gram-negative bacterial lipopolysaccharide (LPS) in the pathogenesis of sepsis was recognized in the 1960's and 70's [2]; therefore, it is not surprising that initial attention to adjunctive treatment measures focused on this molecule. Elucidation of the structure of LPS revealed that

the lipid-A portion was highly conserved among species of Enterobacteriaceae and that the core regions also had considerable conservation. As a result, it was hypothesized that antibodies against these conserved LPS structures might provide protection against a broad range of Gram-negative bacteria. Investigators developed bacterial strains in which the core region of LPS was available to the immune system (i.e. not shielded by O antigen, for example. S. minnesota Re595 [Re chemotype] and Escherichia coli O111:B4, J5 mutant [Rc chemotype]) [3,4]. Pre-clinical work with anti-core LPS antibodies induced by these killed bacterial strains were effective in animal models of sepsis [5,6]. In this manuscript we shall briefly review earlier studies with anti-endotoxin antibodies, and then describe our own studies with a detoxified J5 LPS (dLPS)/group B meningococcal outer membrane complex (OMP) vaccine that progressed to a phase I study in human subjects.

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## 1.1. Early studies with anti-endotoxin antibodies

Based on these earlier studies, Braude and colleagues prepared a whole bacterial vaccine by boiling E. coli O111:B4, Rc chemotype (hereafter, J5 mutant) and raised immune sera in healthy volunteers. In a multi-center clinical trial, patients with suspected Gram-negative bacterial sepsis were given either pre or post-immune sera in addition to standard therapy [7]. Patients with Gram-negative bacteremia who received post-immune sera had a better survival rate (22/91 [24%]) than those receiving pre-immune sera (30/100 [38%] P = 0.041). Among those with either hypotension or in profound shock, there were even more significant differences in favor of the post-immune sera. Despite the therapeutic benefit, there was no evidence that the antisera prevented infection. In this trial investigators were unable to determine whether the antibody fraction of sera was responsible for the improved survival. Further, the antigen in the whole bacterial vaccine responsible for inducing the protection was not clearly identified. Finally, since the "therapeutic product" was material from an individual volunteer and not a reproducibly made reagent, this clinical study must be viewed as a proof of principle rather than the testing of a potential therapeutic product.

Subsequent investigators were unable to confirm the findings of Ziegler et al.; however, none of these studies were similar in design to the original study and none clearly documented the maintenance of anti-endotoxin antibodies (Table 1). In one study, children with meningococcal purpura fulminans were given J5 plasma at the onset of illness [8]. There was no evidence of benefit; however, there was no increase in anti-J5 LPS antibody when measured at 6 h after infusion. In another study, use of J5 plasma was ineffective when given as prophylaxis to surgical patients. This study confirmed the earlier findings of Ziegler et al. [7] that J5 serum did not prevent the development of Gram-negative infection [9]. Similarly, in another clinical trial IgG was prepared from the plasma of volunteers who were immunized with the whole bacterial J5 vaccine [10]. A single infusion of IVIG was ineffective in a clinical trial of patients with sepsis; however, there appeared to be only a two-fold response in anti-J5 LPS antibody in the starting material before fractionation into IVIG. Thus, although the level of anti-core LPS antibodies after infusion was not measured in these patients, it is unlikely that adequate levels of anti-J5 IgG were administered. In yet another study, plasma from blood donors was screened for high levels of naturally occurring anti-core LPS (*S. minnesota*, Re 595) antibody and high titered material was pooled and made into an IVIG [11]. This preparation was compared to standard IVIG in its ability to prevent the onset of sepsis when given as prphylaxis to patients who underwent surgery. In the absence of documented infection, the levels of antibody at 2 days was <50% that of levels obtained at 2 h post infusion [11]. This enriched anti-core LPS IVIG was unable to prevent infection, sepsis or death. Thus, in all of these studies it is likely that inadequate amounts of antibodies were given or inadequate levels of antibody were maintained to test the hypothesis that anti-endotoxin antibodies were effective in the treatment of sepsis.

A number of studies [12–14] have clearly established a relationship between the level of anti-core LPS antibody at the onset of sepsis and outcome. More importantly, a decrease in anti-core LPS antibody during a septic episode forebode a poor outcome [13,15–17]. Consequently, in the absence of documentation that there was an adequate level of circulating anti-endotoxin antibodies, it is difficult to exclude the hypothesis that anti-endotoxin antibodies might be an effective adjunctive therapy for sepsis. Indeed, in small studies, both Schedel and [16] and Fomsgaard [17] and co-workers each demonstrated that maintenance of "adequate levels" of anti-CGL antibody with multiple infusions corresponded to a decrease in circulating endotoxin levels and increased survival.

Despite the fact that early studies with antisera to lipid-A were unsuccessful in treating sepsis in animal models [18], nevertheless, monoclonal antibodies to lipid-A were developed and tested in clinical trials without success [19,20]. Given the repeated failures of anti-core LPS and anti-lipid-A antibodies to affect the outcome of sepsis in clinical trials, subsequent efforts were directed towards the rapidly developing field of cytokine modulation.

## 1.2. Additional therapeutic strategies

Recognition of the important role of  $TNF\alpha$  and IL1 in the development sepsis resulted in multiple clinical trials in which inhibitors of TNF and IL1 activity were tested for therapeutic efficacy in sepsis. After many trials with these and other endogenous mediators of sepsis, no convincing therapeutic effect was detected [21]. In contrast to studies

Table 1
Passive administration of anti-core LPS antibodies for sepsis: previous clinical studies

Study	Product	Number of patients	Ab levels	Outcome
[7]	J5 serum	91	Increased	Reduced mortality, esp if shock
[9]	J5 Plasma	126	Not done	9/136 controls vs. 2/126 patients died
[8]	J5 plasma	40	No increase	No protection in meningococcemia
[11]	Screened IVIG	108	Consumption	No protection
[10]	J5 IVIG	30	Not done	No effect
[16]	"Enriched" IVIG	27	Consumption	Titer-related protection 1/27 vs. 9/28 survival
[17]	Screened IVIG	9	Consumption	Anti-LPS IgG reduced TNF

with anti-endotoxin antibodies which targets an invading pathogen, however, administration of active cytokine antagonist often was associated with increases in lethal infections. These unforeseen adverse events illustrate the difficulty in trying to "fine-tune" the levels of endogenous mediators of sepsis in the host as opposed to efforts to target microbial initiators of sepsis. In view of the difficulties in trying to monitor the effect of therapy on host-defenses as well as the success of the initial clinical trial with J5 antiserum, we decided to reexamine the potential utility of anti-core endotoxin antibodies, such as the J5 antibody. This effort was facilitated by the development of a neutropenic rat model of sepsis in which animals developed a lethal bacterial infection following the administration of relatively low doses of opportunistic pathogens [22].

## 2. Current studies with anti-J5 antibody

We obtained the E. coli O111:J5 strain from Dr. Ziegler and prepared a heat-killed whole bacterial vaccine according to the original method. Antisera raised in rabbits with this vaccine was highly protective in a neutropenic rat model of sepsis, when given at the onset of fever [23] (i.e. as therapy). The effect was clearly dose-related [23], which lent credence to the argument that previous clinical trials with anti-endotoxin antibodies may not have been successful because of inadequate levels of serum administered. We further showed that IgG was the protective fraction in serum and was directed against the core J5 LPS in the whole bacterial vaccine [23]. Six of 8 animals that received affinity purified J5 LPS-specific IgG were protected against lethal Pseudomonas sepsis versus none of 25 animals receiving pre-immune IgG. Importantly, the protection was clearly dose-related with animals receiving 9 ml/kg IgG protected versus none receiving <6 ml/kg [23].

Based on these findings we made a J5 LPS vaccine which was detoxified by removing the ester-linked fatty acids through alkaline treatment [24]. The LPS was not immunogenic when given alone, with alum, with QS21 or when conjugated to tetanus toxoid. When complexed non-covalently with the outer membrane protein of group B-meningococcus, however, the formulation was highly immunogenic in mice, rabbits and rats. Antisera raised with this vaccine was highly protective in a neutropenic rat model after challenge with either Klebsiella or Pseudomonas when the antibody was given either as passive therapy at the time of sepsis, or when antibodies were actively induced by immunization before the start of sepsis. In the latter instance, immunization with this vaccine did not prevent bacteremia, but did reduce mortality. Receipt of anti-J5 antibody reduced circulating levels of endotoxin at 24 h after infusion and reduced the circulating TNF levels compared to the effect with pre-immune sera [24]. Active immunization with the J5dLPS/OMP vaccine promoted the uptake of bacteria from the circulation and killing (i.e. decreased organ bacterial load). Immunization both actively and passively was also protective in another animal model of sepsis, cecal ligation/puncture in mice. This model differs from the neutropenic rat model in that the sepsis is polymicrobial. With these findings we prepared a vaccine for human use.

## 2.1. Phase I clinical study

A Phase I study [25] was conducted in 24 healthy subjects. Subjects received either 5, 10, or 25 µg of vaccine (based on LPS content) at time 0, 1 and 2 months (i.e three total doses). There were few systemic responses (headaches/fever/fatigue) (Table 2). No temperatures >99.9 °F was recorded. Most individuals had a mild-to-moderate degree of tenderness at the injection site, which usually resolved by 48 h. For comparison, the only study to report the incidence of adverse effects with the heat-killed J5 vaccine observed 7/16 incidence of systemic reactions to the initial vaccine, and 3/9 subjects who returned for a second dose [26]. No abnormalities were seen in renal (creatinine, urinalysis), liver (serum alkaline phosphatase, transaminases, bilirubin) or hematologic (leucopenia, anemia) studies compared to baseline studies (data not shown).

Antibody responses were measured by ELISA (Table 3). Compared to pre-immunization levels there was a mean three-fold increase in IgG and IgM levels in the  $10\,\mu g$  group. Five micrograms and  $25\,\mu g$  dosage groups had slightly lower responses. Subjects in all groups had higher baseline levels of IgM antibody to core LPS. We did not assess the affinity of the pre- versus post-immune anti-core

Table 2 Local and systemic reactions following immunization with dJ5 LPS/OMP vaccine

Reactions	Dose (based on dLPS)					
	5 μg	10 μg	25 μg			
Local						
Erythema	$2^{a}$	1	3			
Induration	2	0	4			
Swelling	2	8	6			
Pain						
Severe	0	0	0			
Moderate	8/1 <sup>b</sup>	7/0	12/0			
Mild	10/5	12/5	9/8			
None	6/18	5/19	3/16			
Analgesia	2	2	2			
Systemic						
Fever	1	1	1			
Headache	2	1	0			
Fatigue	0	0	0			
Hematologic						
Anemia	0	1	0			
Leukopenia	0	0	0			

Volunteers were immunized at days 0, 28 and 56 with the indicated dose.

<sup>a</sup> Number of reactions per 24 total immunizations (eight subjects, three doses).

<sup>&</sup>lt;sup>b</sup> Number of reactions at day 1/day 2 after immunization.

Table 3
Anti-J5 LPS ELISA titers of sera from volunteers in the phase I trial

	IgG			IgA			IgM		
$Group^a$	Pre	Post	Fold rise	Pre	Post	Fold rise	Pre	Post	Fold rise
5 (μg)	$1.7^{\rm b}\pm0.28$	$3.6 \pm 0.71$	$2.0 \pm 0.18$	$1.3 \pm 0.14$	$2.6 \pm 0.3$	$2.1 \pm 0.3$	$11.2 \pm 0.9$	$16.9 \pm 1.3$	$1.5 \pm 0.1$
10 (μg)	$2.8 \pm 0.50$	$5.8 \pm 1.9$	$3.3 \pm 0.4$	$4.4 \pm 0.6$	$9.1 \pm 2.0$	$2.0 \pm 0.3$	$18.9 \pm 4.8$	$66.2 \pm 24.0$	$3.2 \pm 1.0$
25 (μg)	$2.1 \pm 0.18$	$4.9 \pm 0.6$	$2.3 \pm 0.3$	$1.8 \pm 0.3$	$3.9 \pm 0.9$	$2.2 \pm 0.5$	$6.5 \pm 1.1$	$18.2 \pm 5.4$	$2.9 \pm 0.6$

<sup>&</sup>lt;sup>a</sup> Eight volunteers in each group received J5 dLPS/OMP vaccine at time 0, days 28 and 56.

LPS antibodies. Six subjects (three high and three low responders) received a single booster dose of  $25\,\mu g$  of vaccine at 12 months to see if it were possible to convert non-responders and to boost the level of responders. High responders were defined as having >2.5-fold increase in serum IgG over baseline, while low responders had <2-fold increase. At 12 months, among responders, pre-boost levels of antibody were still elevated but had decreased by approximately 50%. There was no increase in antibody levels among the high responders following the booster dose. Subjects who did not respond after the primary series did not convert with the booster dose. Plasma from all six subjects was obtained one week after the booster dose. These were evaluated in functional assays.

# 2.2. Functional studies of anti-J5 LPS antibody

With most other vaccines there is usually one functional assay recognized as corresponding to vaccine efficacy. For example, opsonic antibody assays for pneumococcal immunization are thought to better reflect vaccine efficacy than ELISA [27,28]. Viral neutralization assays or serum bactericidal tests have also been thought to correspond to efficacy for other vaccines. In the case of an anti-endotoxin vaccine, however, it is not readily apparent what functional assay would best reflect vaccine efficacy. Many functional activities are initiated by LPS, including induction of cytokines, fever, coagulation as well as the initiation of complement cascades, among a great many other activities. We tested the plasma of the six volunteers in the Phase I study (three high and three low responders) who received a booster dose (25 µg) of vaccine at 1 year in an ex vivo cytokine assay. In this assay, LPS is added to heparinized whole blood and incubated at 37 °C for 24 h [29]. Cytokine generation was then measured in the supernatant. When LPS was pre-mixed with post-immune plasma before addition to the blood, there was a highly significant decrease in TNF (Fig. 1) and in IL6 generation (data not shown) compared to LPS that was exposed to control plasma [25]. This was observed for both low and high responders. When plasma was diluted, however, the higher titered plasma had more activity. Consequently, although the ELISA antibody level

did not correlate with functional activity, those with higher antibody levels did appear to have a higher LPS neutralizing capacity. In a preliminary study, the plasma from a high responder enhanced the clearance of bacteria and endotoxin from the circulation of rats [25].

In yet another functional assay of LPS activity, pre-incubation of human neutrophils with LPS primes the ability to generate superoxide in response to a neutrophil agonist, formyl-methionyl-leucyl-phenylalanine (fMLP) [30]. Pre-incubation of LPS with post-immune sera from three different rabbits (anti-J5-1-3) immunized with the J5dLPS/OMP vaccine reduced the ability of LPS to prime this response (Table 4). Although there did not appear to be an antibody dose-related inhibition of LPS priming based on ELISA antibody levels, we did not dilute out the antisera. When this was done in the ex vivo cytokine induction assay, differences were observed [25]. Based on these initial studies we plan to compare the ability of high and low responder plasma to protect in the cecal ligation puncture and neutropenic rat models of sepsis, to recognize heterologous LPS in other binding assays (fluid phase, and

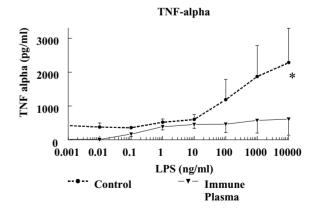


Fig. 1. Effect of pre-incubation of LPS with either post-immunization plasma or control pooled plasma from non-immunized individuals. Different doses of  $E.\ coli$  LPS were added to plasma from either one subject with >3-fold increase in anti-J5 dLPS antibody levels or to control plasma. Then the mixture was added to heparinized whole blood from a J5 LPS-naïve donor. The blood was incubated for 24 h and the supernatant analyzed for TNF $\alpha$ .

<sup>&</sup>lt;sup>b</sup> Serum antibody levels were measured according to our previously described methods [25]. Data represent mean  $\pm$  S.E.M. optical density units (ODU). ODU are defined as the product of the optical density and reciprocal titer for the serum dilution that gives an optical density closest to but still below 1.00 (e.g. OD 0.400 at 1:100 dilution = 40 ODU). Post levels are from the peak antibody level measured on specimens obtained up to 3 months after immunization. Fold rises were calculated for each subject and a geometric mean-fold rise for each group then determined.

Table 4
Post-immune rabbit sera block LPS-primed superoxide response of human neutrophils

J5 LPS
ng/ml)
pplicable
pplicable

Human PMNs were suspended in HBSS/2% human serum and incubated for 60 min at 37  $^{\circ}$ C in medium, medium and LPS or rabbit serum with LPS. The serum from three different rabbits (anti-J5-1; anti-J5-2 and anti-J5-3) immunized, or from non-immunized rabbits (NRS) were used (Anti-J5 LPS antibody levels for each rabbit are indicated in the last column). After washing, the PMNs were stimulated with FMLP ( $10^{-7}$  M) for 10 min in the presence and absence of superoxide dismutase and the change in ferricytochrome C reduction between 0 and 10 min samples determined by absorption at 550 nm. NRS: normal rabbit serum; HBSS: Hank's balanced salt solution. Each condition performed in triplicate. Representative experiment shown of three with similar results.

binding to whole bacteria by flow cytometry) and to neutralize the ability of LPS to induce cytokines by THP1 and RAW cells in vitro. These studies may provide data as to which functional assay may correlate best with protection in animal model of sepsis. This becomes an even more important consideration since there has been considerable and ongoing debate on the methodology for measuring anti-LPS antibodies by ELISA [31].

## 3. Proposed use of anti-endotoxin vaccine

If an effective anti-endotoxin vaccine were available for the prevention and/or treatment of sepsis, then it might be used in several different conditions. Several populations are at higher risk of sepsis and might be considered for immunization: soldiers, police, firefighters, as well as patients undergoing complicated abdominal or genitourinary surgery. Routine immunization of the first three groups would require that the antibody response be long-lived. In our phase I study, subjects with elevated anti-J5 LPS antibody responses after initial immunization still had elevated antibodies at 12 months [25]. In the case of patients undergoing elective surgery, an effective anti-endotoxin vaccine would need to induce antibodies after one or two doses of vaccine. Co-administration of the vaccine with an adjuvant might accelerate the antibody response in a manner similar to that of the oligonucleotide, CpG, given with hepatitis B (a vaccine also given in three doses) [32]. Since after acute injury there is a Th2 polarization, patients admitted with burns or trauma might respond to active immunization [33,34]. We administered experimental Klebsiella and Pseudomonas vaccines to patients admitted following severe trauma and found that they responded well to both vaccines [35].

Alternatively, anti-core LPS antibodies could be given passively to septic patients. In this instance, it would be essential to monitor the circulating levels of anti-core LPS antibodies. In our own pre-clinical studies in neutropenic rats there was a clear dose-related protection [23], and previous clinical trials did not pay adequate attention to the maintenance of antibody levels. Additional doses of antibody may be required during a septic episode. In patients who become septic despite active immunization with an anti-endotoxin vaccine, supplementation with passive administration of antibodies may be required to counter any consumption of antibody, as was documented in previous trials.

#### 4. Conclusions

Our own bias is that many of these previous studies that investigated the efficacy of anti-endotoxin antibody therapy did not adequately measure the amount of antibody administered and did not insure adequate levels of antibody after initial infusions. Consequently, the potential role of anti-core endotoxin antibody therapy has not been sufficiently tested to discard the hypothesis. In monitoring the adequacy of therapy, the discrepancy between the ELISA antibody levels in human subjects and their activity in functional studies needs to be confirmed in a more rigorous fashion. Given the number of functional assays with which one might measure anti-endotoxin activity, this may become a daunting task. The conflicting data with previous studies of anti-endotoxin antibody therapy demands, however, that this effort be pursued in order to better evaluate the response to vaccine such as the one under present study. The current studies suggest that monitoring responses with functionally relevant assays may be an important component of clinical trials with anti-endotoxin antibodies. Moreover, our earlier studies in a neutropenic rat model of sepsis demonstrated the importance of giving adequate levels of anti-endotoxin antibodies [23]. The more recent study in human subjects found that even though the plasma from both high and low responders neutralized the cytokine-inducing activity of LPS, nevertheless, the activity was greater for the high responders [25]. Consequently, it may be desirable to devise strategies to improve the antibody response with this J5 dLPS/OMP complex vaccine.

Future studies will be directed toward administration of this vaccine with adjuvants that may boost the level of antiendotoxin antibodies and enhance the functional activity of the preparation. These strategies are currently being investigated.

#### References

 Martin GS, Mannino DM, Eaton S, Moss M. The epidemiology of sepsis in the United States from 1979 through 2000. N Engl J Med 2003;348:1546–54.

- [2] Braude AE, Jones JL, Douglas H. The behavior *Escherichia coli* endotoxin (somatic antigen) during infectious arthritis. J Immunol 1960:90:297–312
- [3] McCabe WR. Immunization with R mutants of S. minnesota. Part I. Protection against challenge with heterologous Gram-negative bacilli. J Immunol 1972;108:601–10.
- [4] Ziegler EJ, Douglas H, Sherman JE, Davis CE, Braude AI. Treatment of E. coli and Klebsiella bacteremia in agranulocytic animals with antiserum to a UDP-GAL epimerase-deficient mutant. J Immunol 1973:111:433–8
- [5] Ziegler EJ, McCutchan JA, Douglas H, Braude AI. Prevention of lethal *Pseudomonas* bacteremia with epimerase-deficient *E. coli* antiserum. Trans Assoc Am Physicians 1975;88:101–8.
- [6] Johns M, Skehill A, McCabe WR. Immunization with rough mutants of *Salmonella minnesota*. IV. Protection by antisera to O and rough antigens against endotoxin. J Infect Dis 1983;147:57–62.
- [7] Ziegler EJ, McCutchan JA, Fierer J, Glauser MP, Sadoff JC, Douglas H, et al. Treatment of Gram negative bacteremia and shock with human antiserum to a mutant *Escherichia coli*. N Engl J Med 1982;307:1225–30.
- [8] J5 Group. Treatment of severe infectious purpura in children with human plasma from donors immunized with *Escherichia coli* J5: a prospective double-blind study. J Infect Dis 1992;165:695–701.
- [9] Baumgartner JD, Glauser MP, McCutchan JA, Ziegler EJ, Van Melle G, Klauber MR, Vogt M, Muehlen E, Luethy R, Chiolero R, Geroulanos S. Prevention of Gram-negative shock and death in surgical patients by antibody to endotoxin core glycolipid. Lancet 1985;2:59–63.
- [10] Calandra T, Glauser MP, Schellekens J, Verhoef J. Swiss-Dutch J5 immunoglobulin study group. Treat of Gram-negative septic shock human IgG antibody to *Escherichia coli* J5: a prospective, double-blind, randomized trial. J Infect Dis 1988;158:312–9.
- [11] The Intravenous Immunoglobulin Collaborative Study Group. Prophlactic intravenous immunoglobulin administration of standard immunoglobulin as compared with core-lipopolysaccharide immune globulin in patients at high risk of postsurgical infection. N Engl J Med 1992;327:234–40.
- [12] Pollack M, Huang AI, Prescott RK, Young LS, Hunter KW, Cruess DF, et al. Enhanced survival in *Pseudomonas aeruginosa* septicemia associated with high levels of circulating antibody to *Escherichia coli* endotoxin core. J Clin Invest 1983;72:1874–81.
- [13] Goldie AS, Fearon KCH, Ross JA, Barclay R, Jackson RE, Grant IS, Ramsay G, Blyth AS, Howie JC. Natural cytokine antagonists and endogenous antiendotoxin core antibodies in sepsis syndrome. JAMA 1995;274:172–7.
- [14] Zinner SH, McCabe WR. Effects of IgM and IgG antibody in patients with bacteremia due to Gram-negative bacilli. J Infect Dis 1976:133:37–45
- [15] Nys M, Damas P, Joassin L, Lamy M. Sequential anti-core glycolipid immunoglobulin antibody activities in patients with and without septic shock and their relation to outcome. Ann Surg 1993;217:300–6.
- [16] Schedel I, Dreikhausen U, Nentwig B, Hockenschneider M, Rauthmann D, Balikcioglu S, Coldeway R, Deicher H. Treatment of Gram-negative septic shock with an immunoglobulin preparation: a prospective, randomized clinical trial. Crit Care Med 1991;9:1104– 13.
- [17] Fomsgaard A, Baek L, Fomsgaard JS, Engquist A. Preliminary study on treatment of septic shock patients with antilipopolysaccharide IgG from blood donors. Scand J Infect Dis 1989;21:697–708.
- [18] Bruins SC, Stumacher R, Johns MA, McCabe WR. Immunization with R mutants of *Salmonella minnesota*. III. Comparison of the protective effect of immunization with lipid A and the Re mutant. Infect Immun 1977;17:16–20.
- [19] Ziegler EJ, Fisher Jr CJ, Sprung CL, Straube RC, Sadoff JC, Foulke GE, et al. Treatment of Gram-negative bacteremia in septic shock with HA-1A human monoclonal antibody against endotoxin: a randomized, double-blind, placebo-controlled trial. N Engl J Med 1991;325:429–36.

- [20] Greenman RL, Schein RMH, Martin MA, Wenzel RP, Mac Intyre NR, Emmanuel G, Chmel H, Kohler RB, McCarthy M, Plouffe J, Russell JA. A controlled clinical trial of E5 murine monoclonal IgM antibody to endotoxin in the treatment of Gram-negative sepsis. JAMA 1991:266:1097–102.
- [21] Zeni F, Freeman B, Natanson C. Anti-inflammatory therapies to treeat sepsis and septic shock: a reassessment. Crit Care Med 1997;25:1095–100.
- [22] Collins HH, Cross AS, Dobek A, Opal SM, McClain JB, Sadoff JC. Oral ciprofloxacin and anti-lipopolysaccharide monoclonal antibody protect leukopenic rats from lethal infection. J Infect Dis 1989:159:1073–82
- [23] Bhattacharjee AK, Opal SM, Palardy JE, Drabick JJ, Collins H, Taylor R, Cotton A, Cross AS. Affinity-purified *Escherichia coli* J5 lipopolysaccharide-specific IgG protects neutropenic rats against Gram-negative bacterial sepsis. J Infect Dis 1994;170:622–9.
- [24] Bhattacharjee AK, Opal SM, Taylor R, Naso R, Semenuk M, Zollinger WD, Moran EE, Young L, Hammack C, Sadoff JC, Cross AS. A noncovalent complex vaccine prepared with detoxified *Escherichia coli* J5 (Rc Chemotype) lipopolysaccharide and *Neisseria meningitidis* Group B outer membrane protein produces protective antibodies against Gram-negative bacteremia. J Infect Dis 1996;173:1157–63.
- [25] Cross AS, Opal SM, Palardy JE, Drabick JJ, Warren HS, Huber C, Cook P, Bhattacharjee AK. Phase I study of detoxified *Escherichia coli* J5 lipopolysaccharide (J5dLPS)/group B meningococcal outer membrane protein (OMP) complex vaccine in human subjects. Vaccine 2003;21:4576–88.
- [26] Schwartzer TA, Alcid DV, Numsuwan V, Gocke DJ. Characterization of the human anti-body response to an *Escherichia coli* O111:B4 (J5) vaccine. J Infect Dis 1988;158:1135–6.
- [27] Johnson SE, Rubin L, Romero-Steiner S, Dykes JK, Pais LB, Rizvi A, Ades E, Carlone GM. Correlation of opsonophagocytosis and passive protection assay using human anticapsular antibodies in an infant mouse model of bacteremia for Streptococcus pneumoniae. J Infect Dis 1999;180:133–40.
- [28] Kim KH, Seoh JU. Evaluation of antibody responses to pneumococcal vaccines with ELISA and opsonophagocytosis assay. J Korean Med Sci 1999;14:475–9.
- [29] Kovach NL, Yee E, Munford RS, Raetz CR, Harlan JM. Lipid IVA inhibits synthesis and release of tumor necrosis factor induced by lipopolysaccharide in human whole blood ex vivo. J Exp Med 1990:172:77–84
- [30] Guthrie LA, McPhail LC, Henson PM, Johnston Jr RB. Priming of neutrophils for enhanced release of oxygen metabolites by bacterial lipopolysaccharide. Evidence for increased activity of the superoxide-producing enzyme. J Exp Med 1984;160:1656–71.
- [31] Warren HS, Amato SF, Fitting C, Black KM, Coiselle PM, Pasternack MS, et al. Assessment of ability of murine and human anti-lipid A monoclonal antibodies to bind and neutralize lipopolysaccharide. J Exp Med 1993;177:89–97.
- [32] Davis HL, Cooper CL, Morris ML, Elfer SM, Cameron DW, Heathcote J, CpG ODN is safe and highly effective in humans as an adjuvant in hepatitis B vaccines. Preliminary results of phase I trial with CpG 7909. In: Proceedings of the Third Annual Conference on Vaccines (Abstract S25); 2000.
- [33] Lyons A, Kelly JL, Rodrick ML, Mannick JA, Lederer JA. Major injury induces increased production of interleukin-10 by cells of the immune system with a negative impact on resistance to infection. Ann Surg 1997;226:450–60.
- [34] Ginnoudis PV, Smith RM, Banks RE, Windsor ACJ, Dickson RA, Guillou PJ. Stimulation of inflammatory markers after blunt trauma. Br J Surg 1998;85:986–90.
- [35] Campbell WN, Hendrix E, Cryz SJ, Cross AS. Immunogenicity of a 24-valent Klebsiella capsular polysaccharide vaccine and an 8-valent Pseudomnas O-polysaccharide conjugate vaccine administered to acute trauma victims. Clin Infect Dis 1996;23:179–81.